



Discovery and optimization of RO-85, a novel drug-like, potent, and selective P2X₃ receptor antagonist

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ARTICLE INFO

Article history:

Received 24 September 2009

Revised 6 December 2009

Accepted 10 December 2009

Available online 16 December 2009

Keywords:

P2X₃

Ion channel

RO-85

Inflammatory pain

Neuropathic pain

Drug-like

P2X₃ receptor antagonist

ABSTRACT

Despite the extensive literature describing the role of the ATP-gated P2X₃ receptors in a variety of physiological processes the potential of antagonists as therapeutic agents has been limited by the lack of drug-like selective molecules. In this paper we report the discovery and optimization of RO-85, a novel drug-like, potent and selective P2X₃ antagonist. High-throughput screening of the Roche compound collection identified a small hit series of heterocyclic amides from a large parallel synthesis library. Rapid optimization, facilitated by high-throughput synthesis, focusing on increasing potency and improving drug-likeness resulted in the discovery of RO-85.

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P2X receptors are ligand gated ion channels activated by adenosine 5'-triphosphate (ATP) and related di- and tri-phosphate nucleotides. Seven P2X receptor subunits have been identified P2X₁₋₇ and each channel shown to be assembled from three subunits.¹ Understanding the pharmacology of the channels is complex in that, with the exception of P2X₆, in addition to homomeric channels, each subunit also has the ability to form heteromeric channels and a total of seven heteromeric P2X family members have been identified.² Homomeric P2X₃, and the closely related heteromultimeric P2X_{2/3}, receptors are predominantly colocalized on small to medium diameter sensory afferent neurons and have become increasingly recognized as playing a major role in mediating the primary sensory effects of ATP. P2X₃-KO mice demonstrate a reduced sensitivity to thermal stimuli and decreased pain behaviors.³ Furthermore reduction of P2X₃ expression through intrathecal administration of P2X₃-selective antisense,⁴ and more recently siRNA,⁵ also causes a significant decrease in pain behaviors in mice. By eliminating the P2X₃ subunit these gene deletion and knockdown techniques also eliminate the P2X_{2/3} receptor so a clear understanding of the receptor responsible for these effects is hard to ascertain.

Despite significant advances in elucidating the pharmacology of P2X₃ receptors, the field has suffered from a lack of drug-like, potent and selective ligands. Many of the reported selective P2X₃ antagonists (Fig. 1), including the most recently reported highly potent peptide antagonist Spinorphin (**1**),⁶ either do not report selectivity over P2X_{2/3} or are dual P2X₃/P2X_{2/3} antagonists like TNP-ATP (**2**)⁷ and A-317491 (**3**),⁸ as well as our recently published diaminopyrimidines, (**4a**) and (**4b**).^{9,10}

In addition to the unknown, or lack of selectivity over P2X_{2/3} making their pharmacology hard to understand, these ligands also show poor drug-like properties. TNP-ATP and A-317491, similar to many of the non-selective P2X₃ receptor antagonists, contain polyacidic functional groups that limit their bioavailability and overall potential as drug candidates. Despite this A-317491 has been shown to be active in a number of in vivo models of chronic inflammatory pain, neuropathic pain⁸ and overactive bladder.¹¹ No data has been published on the drug-like properties of spinorphin which has also been shown to be effective in the tail pinch assay following intracerebroventricular (ICV) administration.¹² In this Letter we report the discovery and optimization of a series of drug-like potent and selective P2X₃ antagonists.

A high-throughput screening campaign of the Roche compound collection using the rat recombinant P2X₃ receptor was performed. Disappointingly, in addition to a very overall low hit rate of 0.01%, many of the more potent sub-micromolar hits contained similar

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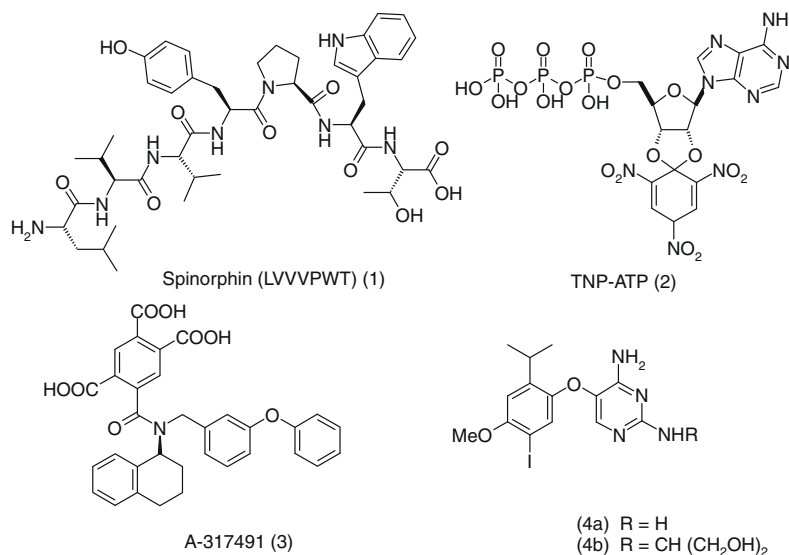


Figure 1. Structures of known P2X₃ antagonists.

poly-acidic functionality to the known antagonists with their implied poor drug-like properties. These were not deemed suitable starting points for a medicinal chemistry program aimed at producing potential therapeutic agents so a careful analysis of the less potent hits was undertaken. Four compounds, initially synthesized as part of a ~4500 member parallel synthesis library, were identified as hits. The library was based on a 2,4-disubstituted-oxazole-5-carboxylic acid amide core structure. The hits were characterized by a 2-phenyl substituent on the oxazole core and, more specifically, the presence of a 1-methyl-2-(4-pyrimidin-2-yl-piperazin-1-yl)-ethylamine amide (Fig. 2).

A detailed pharmacological and drug-likeness characterization of (5a) was undertaken. (5a) was shown to be a selective antagonist of rP2X₃ (pIC₅₀ 6.6) with no antagonist activity at P2X_{2/3} or other family members tested (pIC₅₀ <5). In a standard CEREP profile of 80 other enzymes, receptors and ion channels only five other activities were identified in the 5–10 μM range.¹³ In an assessment of drug-likeness; with positive attributes of moderate molecular weight (434) and calculated log *P* (2.7), and high permeability without efflux it was disappointing to find high in vitro clearance in rat and human microsomes and hepatocytes, and >99% protein binding in rat plasma. In addition (5a) was a moderate inhibitor of the CYP450 isozymes 3A4 and 2D6 with IC₅₀'s <10 μM. A parallel optimization approach exploring replacement of the oxazole core and its substituents, and the amide portion of the molecule was undertaken.

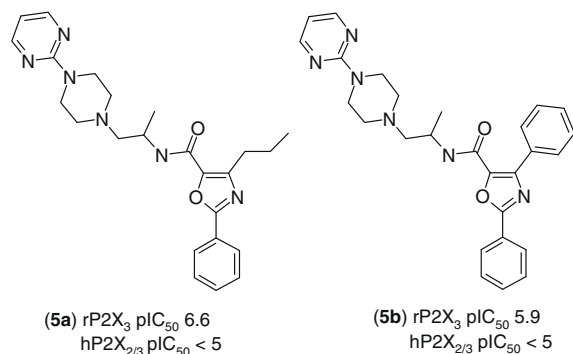


Figure 2. Hits structures identified by high-throughput screening and their pIC₅₀'s at the rat P2X₃ receptor.

The role of the oxazole scaffold was investigated using both a general collection of acids fitting a similar motif and by a traditional iterative SAR approach. A representative set of similar amides is shown in Table 1. For five-membered scaffolds, pyrazole amides (6a) and (6b) and furans (6c) and (6d) had similar activity to one of the other initial hits (5b). Substitutions that altered the geometry of the amide bond, (6g) and (6h) were not tolerated. Substitution of the butyl or phenyl group for a methyl gave a significant drop in activity (6i) versus (5a) and (5b). Interestingly, substitution of the phenyl groups in a 3,4 relationship as in (6j) and (6f) led to no loss in activity.

A six-membered ring scaffold was also accepted, as shown in examples (8a–8f), even with only one aryl substituent. An interesting 5-5-bicyclic core, 1-methyl-3-phenyl-1*H*-thieno[2,3-*c*]pyrazole-5-carboxylic amide (7), discovered using parallel synthesis was found to show promising opportunities by allowing truncation of the propyl group to a methyl group.

Exploration of the piperazinyl-ethylamine linker of the molecule showed little opportunity for modification (see Fig. 3) with the exception of a preference for the *R*-stereochemistry (Table 2). The pyrimidine group could, however be modified with other heterocycles, without losing activity at the P2X₃ receptor (9a–9f) as long as a nitrogen was present in the 2-position to act as an elector acceptor (see Table 2). The *m*-pyridine and *p*-pyridine were inactive (9g and 9h). Interestingly, a simple carbonyl group was shown to be sufficient for good affinity as long as the group remained planar. For example, acetyl amide (10a), thioacetyl amide (10c) and urea (10d) were all active, whereas the cyclopropylcarbonylamide (10f) and other amides not shown here, the substituted urea (10e), sulfonamide (10g) and ethyl carbamate (10h) were inactive. Of particular significance was the promising activity of (10a) with a concurrent improvement in molecular weight and lipophilicity. These analogs were synthesized easily starting from Boc-D-alaninal as shown in Scheme 1.

Reaction of Boc-D-alaninal with *N*-acetylpiperazine in the presence of NaBH(OAc)₃ afforded intermediate (15) in 85% yield. Removal of the Boc group with TFA, followed by coupling with commercially-available 1-methyl-3-phenyl-1*H*-thieno[2,3-*c*]pyrazole-5-carboxylic acid using a HBTU coupling afforded the amide, which was converted the oxalate salt (16).¹⁴ Similar reductive amination conditions, followed by deprotection were used to prepare *N*-arylpiperidines (14a–14h), which were coupled to 2-(4-fluorophenyl)-4-propyl-oxazole-5-carboxylic acid¹⁵ using DCC and HOBT

Table 1
Heterocyclic core SAR

No.	R	pIC ₅₀ ^{a,b,c}
6a		6.0
6b		6.0
6c		6.2
6d		5.7
5b		5.9
6e		5.4
6f		5.7
6g		<5
6h		<5
7		6.7
6i		5.3

Table 1 (continued)

No.	R	pIC ₅₀ ^{a,b,c}
6j		5.9
8a		5.9
8b		5.3
8c		6.4
8d		6.3
8e		6.3
8f		6.9

Ar = *p*-F-phenyl^a FLIPR: mean pIC₅₀, rP2X₃ CHO cell.^b FLIPR: mean pIC₅₀ <5.0, hP2X_{2/3} 1321n1c cells.^c pIC₅₀ values are the mean of at least three experiments performed in triplicates, standard deviation +20%.

to afford the corresponding amides (**9a–9h**). Intermediate (**17**), prepared by reductive amination of Boc-D-alaninal with Cbz-piperazine, was coupled the same acid to provide intermediate (**18**). Removal of the Cbz group by catalytic hydrogenation afforded the free piperazine, which was functionalized to afford (**10a–10f**) using standard conditions.

The thienopyrazole template, discovered with analog (**7**) and which had the advantage of having good drug-likeness and for not requiring a longer alkyl chain with the potential metabolic liabilities, was explored further. The results are shown in Table 3. Substitution of the phenyl ring in the 3-position with an *ortho* fluorine also led to active analog (**11a**), however an *ortho*-methyl substituent was less tolerated (**11b**). Increasing the size of the N-1 substituent to an ethyl slightly decreased activity (**11c**) and introduction of a phenyl group led to an inactive analog (**11d**). Changing the sulfur to an oxygen to give (**12**) led to an inactive molecule.

Combining the substitution of an acetyl for a pyrimidyl group and the thienopyrazole core template led to potent molecules (**11f–11h**, Table 3). Resolution of the amide side chain resulted in the identification of (*R*) as the preferred stereochemistry; thus 1-methyl-3-phenyl-1*H*-thieno[2,3-*c*]pyrazole-5-carboxylic acid [(*R*)-

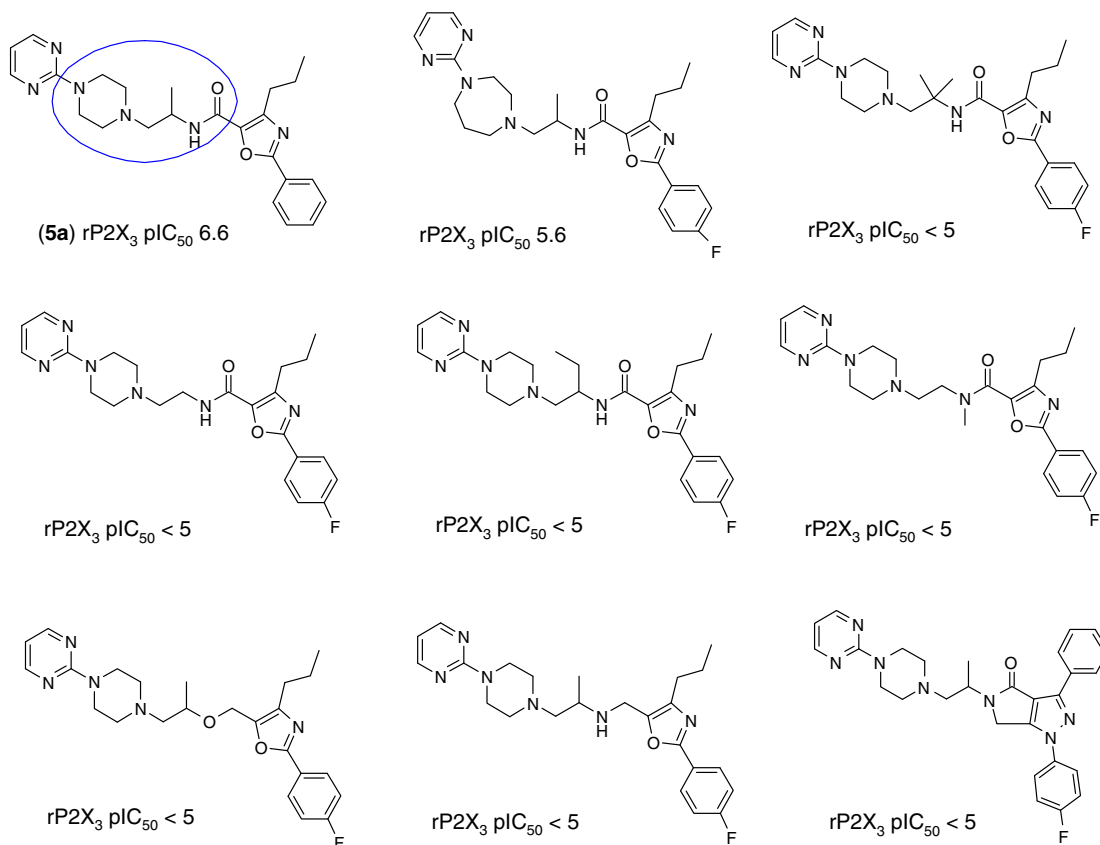


Figure 3. Linker modifications and their pIC₅₀'s at the rat P2X₃ receptor.

Table 2
Acceptor group SAR

No.	R	Stereo-chem.	pIC ₅₀ ^{a,b,c}
9a		R	7.3
9b		S	5.7
9c		R	5.8
9d		R	6.3
9e		R	5.9
9f		R,S	6.2

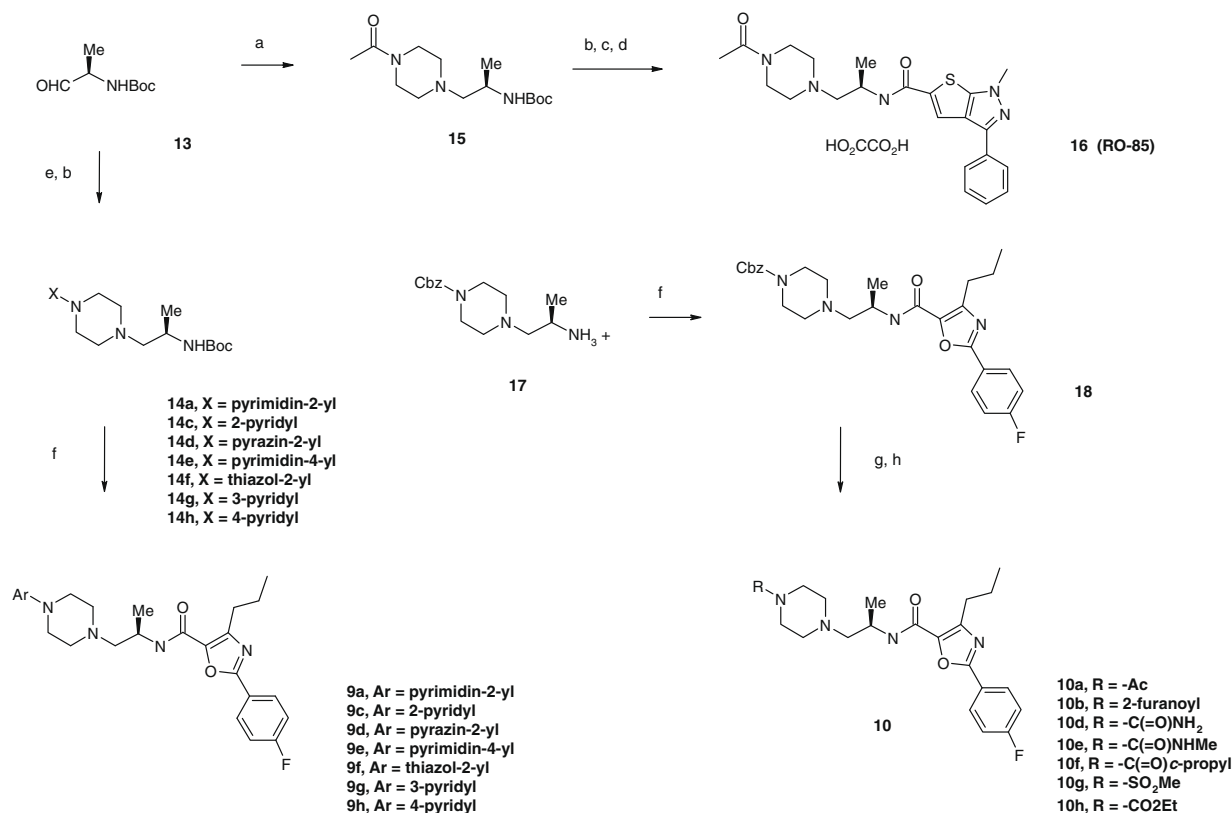
Table 2 (continued)

No.	R	Stereo-chem.	pIC ₅₀ ^{a,b,c}
9g		R,S	<5
9h		R	<5
10a	Ac	R	6.4
10b		R	5.6
10c		R,S	7.0
10d		R	6.0
10e		R	<5
10f		R	<5
10g	–SO ₂ Me	R	<5
10h	–CO ₂ Et	R	<5

^a FLIPR: mean pIC₅₀, rP2X₃ CHO cell.

^b FLIPR: mean pIC₅₀ <5.0, hP2X_{2/3} 1321n1c cells.

^c pIC₅₀ values are the mean of at least three experiments performed in triplicates, standard deviation +20%.



Scheme 1. Reagents and conditions: (a) *N*-acetylpiperidine, 1.2 equiv, DCE, NaBH(OAc)₃, 2 equiv, 20 h; (b) TFA, CH₂Cl₂ 2 h; (c) HBTU, 1.5 equiv, excess DIEA, THF; (d) oxalic acid, 1.1 equiv, ether; (e) CH₂Cl₂; *N*-arylpiperazines (2-piperazin-1-ylpyrimidine, 1-pyridin-2-ylpiperazine, pyrazin-2-ylpiperazine, 4-piperazin-1-ylpyrimidine, 1-thiazol-2-ylpiperazine, 1-pyridin-3-ylpiperazine, or 1-pyridin-4-ylpiperazine), 1.2 equiv, DCE, NaBH(OAc)₃, 2 equiv; (f) (i) 2-(4-fluorophenyl)-4-propyl-oxazole-5-carboxylic acid, DCC on resin, HOBt, CH₂Cl₂, 2 h, (ii) amine (**14a–14h**, **17**), CH₂Cl₂, DIEA 20 h; (g) Pd/C (10% Degussa), H₂, EtOH; (h) for **10b** 2-furoylchloride, CH₂Cl₂, DIEA, for **10d** KOCN, AcOH, H₂O; for **10e** CH₃NCO, CH₂Cl₂; for **10f** c-C₃H₅COCl, CH₂Cl₂, DIEA, for **10g** CH₃SO₂Cl, CH₂Cl₂, DIEA, for **10h** ClCO₂Et, CH₂Cl₂, DIEA.

Table 3
SAR of 1H-thieno[2,3-*c*]pyrazole-5-carboxamides

No.	X	R ¹	R ²	R ³	pIC ₅₀
7	S	Me	Ph	Pyrimidyl	6.7
11a	S	Me	<i>o</i> -F-Ph	Pyrimidyl	7.2
11b	S	Me	<i>o</i> -Me-Ph	Pyrimidyl	5.7
11c	S	Et	Ph	Pyrimidyl	6.3
11d	S	Ph	Ph	Pyrimidyl	<5
11e	S	Ph	Me	Pyrimidyl	5
11f	S	Me	Ph	Acetyl	7.0
11g	S	Me	<i>p</i> -Me-Ph	Acetyl	7.4
11h	S	Me	<i>o</i> -F-Ph	Acetyl	7.4
12	O	Me	Ph	Pyrimidyl	<5

2-(4-acetyl-piperazin-1-yl)-1-methyl-ethyl]-amide (**16**), identified as RO-85 was discovered.

RO-85 is a potent P2X₃ receptor antagonist at the rat and human receptors (rP2X₃ pIC₅₀ = 7.5; hP2X₃ = 6.4) selective over hP2X_{2/3} and the other P2X family members tested (pIC₅₀ < 5). No other pharmacological activity was seen in a standard CEREP panel of 80 other enzymes, receptors and ion channels (no activity >50% at 10 μM).¹³ Favorable molecular properties comparable to the

original hit structure were also retained (MW 425, c log *P* 2.8). RO-85 showed significantly higher in vitro metabolic stability¹⁶ in rat and human liver microsomes with 89% protein binding in rat plasma. No inhibition of the CYP450 isozymes was seen at concentrations <15 μM. When dosed orally to rats RO-85 was 89% orally bioavailable with a half-life of 1.6 h.

In summary we have identified a novel, potent and selective P2X₃ receptor antagonist RO-85. This was discovered through the rapid optimization of a hit series identified by high-throughput screening facilitated by parallel synthesis. RO-85 is exquisitely selective for the P2X₃ receptor subtype over other P2X family members and other pharmacological targets and has suitable pharmacokinetic properties to explore the therapeutic potential of P2X₃ antagonists in vivo. Additional SAR studies will be published in the future.

References and notes

- Gever, J. R.; Cockayne, D. A.; Dillon, M. P.; Burnstock, G.; Ford, A. P. D. W. *Pfluegers Arch.* **2006**, 452, 513.
- Dubyak, G. R. *Mol. Pharmacol.* **2007**, 72, 1402.
- Cockayne, D. A.; Hamilton, S. G.; Zhu, Q.; Dunn, P. M.; Zhong, Y.; Novakovic, S.; Malmberg, A. B.; Cain, G.; Berson, A.; Kassotakis, L.; Hedley, L.; Lachnit, W. G.; Burnstock, G.; McMahon, S. B.; Ford, A. P. D. W. *Nature* **2000**, 407, 1011.
- Barclay, J.; Patel, S.; Dorn, G.; Wotherspoon, G.; Moffatt, S.; Eunson, L.; Abdel'al, S.; Natt, F.; Hall, J.; Winter, J.; Bevan, S.; Wishart, W.; Fox, A.; Ganju, P. J. *Neurosci.* **2002**, 22, 8139.
- Dorn, G.; Patel, S.; Wotherspoon, G.; Hemmings-Mieszczyk, M.; Barclay, J.; Natt, F. J. C.; Martin, P.; Bevan, S.; Fox, A.; Ganju, P.; Wishart, W.; Hall, J. *Nucleic Acids Res.* **2004**, 32, e49/1.
- Jung, K.; Moon, H. D.; Lee, G. E.; Lim, H.; Park, C.; Kim, Y. J. *Med. Chem.* **2007**, 50, 4543.

7. Virginio, C.; Robertson, G.; Surprenant, A.; North, R. A. *Mol. Pharmacol.* **1998**, *53*, 969.
8. Jarvis, M. F.; Burgard, E. C.; McGaraughty, S.; Honore, P.; Lynch, K.; Brennan, T. J.; Subieta, A.; Van Biesen, T.; Cartmell, J.; Bianchi, B.; Niforatos, W.; Kage, K.; Yu, H.; Mikusa, J.; Wismer, C. T.; Zhu, C. Z.; Chu, K.; Lee, C.; Stewart, A. O.; Polakowski, J.; Cox, B. F.; Kowaluk, E.; Williams, M.; Sullivan, J.; Faltynek, C. *Proc. Natl. Acad. Sci. U.S.A.* **2002**, *99*, 17179.
9. Carter, D. S.; Alam, M.; Cai, H.; Dillon, M. P.; Ford, A. P. D.; Gevert, J. R.; Jahangir, A.; Lin, C.; Moore, A. G.; Wagner, P. J.; Zhai, Y. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 1628.
10. Jahangir, A.; Alam, M.; Carter, D. S.; Dillon, M. P.; Du Bois, D. J.; Ford, A. P. D.; Gevert, J. R.; Lin, C.; Wagner, P. J.; Zhai, Y.; Zira, J. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 1632.
11. Ito, K.; Iwami, A.; Katsura, H.; Ikeda, M. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **2008**, *377*, 483.
12. Honda, M.; Okutsu, H.; Matsuura, T.; Miyagi, T.; Yamamoto, Y.; Hazato, T.; Ono, H. *Jpn. J. Pharmacol.* **2001**, *87*, 261.
13. CEREP 80 screen performed by Cerep Corp. 15318 NE 95th St. Redmond, WA 98052, USA. www.cerep.com.
14. The mono-oxalate salt provided a free-flowing, non-hydroscopic salt preferable to other salts prepared, including the TFA salt and HCl salt.
15. Brotherton-Pleiss, C.E.; Dillon, M.P.; Gleason, S.K.; Lin, C.J.J.; Schoenfeld, R.C.; Villa, M.; Zhai, Y. U.S. Patent 2007037974, 2007; *Chem. Abstr.* **2007**, *146*, 229379.
16. HLM = 1.76 $\mu\text{L}/\text{min}/\text{mg}$ (high stability), RLM = 43.7 $\mu\text{L}/\text{min}/\text{mg}$ (medium stability), rat hepatocytes = 3.93 $\text{mL}/\text{min}/\text{million cells}$ (high stability).